

Cyclobutane-Containing Alkaloids: Origin, Synthesis, and Biological Activities

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Abstract: Present review describes research on novel natural cyclobutane-containing alkaloids isolated from terrestrial and marine species. More than 60 biological active compounds have been confirmed to have antimicrobial, antibacterial, antitumor, and other activities. The structures, synthesis, origins, and biological activities of a selection of cyclobutane-containing alkaloids are reviewed. With the computer program PASS some additional biological activities are also predicted, which point toward new possible applications of these compounds. This review emphasizes the role of cyclobutane-containing alkaloids as an important source of leads for drug discovery.

Keywords: Alkaloids, cyclobutane-containing, anticancer, antibacterial, synthesis, activities, terrestrial, marine.

INTRODUCTION

Organic compounds containing four-membered ring(s) represent unusual group of metabolites including natural products and/or drugs. The cyclobutane unit is found as a basic structural element in a wide range of naturally occurring compounds in bacteria, fungi, plants, and marine invertebrates. It is also generated transiently in primary and secondary metabolisms [1]. Many biological activities are showed and may serve as potential drug leads or provide new ideas for the study of enzyme mechanisms, and/or organic synthesis [2]. Some cyclobutane compounds such as amino acids, peptides and nucleosides showed protective properties against UV radiation [3]. In the skin many molecules may absorb UV radiation upon exposure. In particular, cellular DNA strongly absorbs shorter wavelength solar UV radiation, resulting in various types of DNA damage. Among the DNA photoproducts produced the cyclobutane pyrimidine dimers are predominant [4].

Although cyclobutanes have been known for more than a century, their use as synthetic intermediates has only flourished in the last forty years. The structures and syntheses of cyclobutanoid fatty acids, amino acids, mono-, sesqui-, di-, and triterpenes, steroids, and other compounds have recently been reported [2,5,6].

In the present review, we will focus on origin, structures, and biological activities of natural cyclobutane-containing alkaloids and selected related compounds. Their structure and biological activities, modes of action, and future prospects are discussed.

This paper is a short survey of cyclobutane-containing alkaloids that are deemed as naturally occurring. Also, this is

the first article to review natural alkaloids comprising a cyclobutane unit.

TERRESTRIAL CYCLOBUTANE-CONTAINING ALKALOIDS

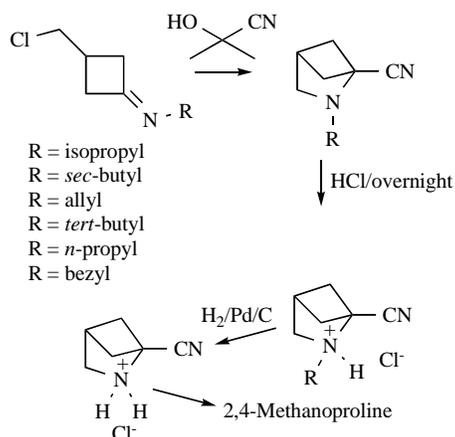
GC-MS was used to analyze volatiles in the headspace of 'Fortress' onion bulbs inoculated with *Fusarium oxysporum*, *Botrytis allii*, *Erwinia carotovora* subsp. *carotovora*, *Aspergillus niger*, or *Penicillium aurantiogriseum*. Among 130 volatile metabolites of 'Fortress' onion bulbs were found a simple ethylcyclobutane and 2-azabicyclo[3.2.0]hept-6-ene (**1**) [7]. 2,4-Methanoproline (2-carboxy-2,4-methanopyrrolidine) (**2**) and 2,4-methanoglutamic acid (1-amino-1,3-dicarboxycyclobutane) were isolated from seeds of *Ateleia herbert smithii* (Leguminosae) [8].

The non-protein amino acids of the legume genus *Bocoa* (Papilionoideae; Swartzieae) were surveyed by LC-MS and GC-MS using extracts of herbarium leaf fragments. 2,4-Methanoproline (**2**) have also been detected in *Ateleia herbert smithii*, *Bocoa alterna*, *B. decipiens*, *B. limae*, *B. mollis*, and *B. racemulosa* [9]. Compound (**2**) was formerly suggested to have insect repellent/antifeedant activity [10].

Synthesis of 2,4-methanoproline and derivatives have been reported by several authors. Most of the syntheses of 2,4-methanoproline are accomplished by an intramolecular light-induced [2+2] cycloaddition of an appropriate diene [11,12]. Only Gaoni used an intramolecular cyclization strategy for his approach [13]. Also analogues containing the 2-azabicyclo[2.1.1]hexane skeleton are often synthesized by the light-induced [2+2] cyclization [14-17]. The rearrangement of an appropriate bromohydrine to synthesize 5-hydroxy-2-azabicyclo[2.1.1]-hexanes has also been described [18,19]. A two-step synthetic approach toward 3-(chloromethyl)cyclobutanone was described and used in the synthesis of 2,4-methanoproline analogues [20]. The key

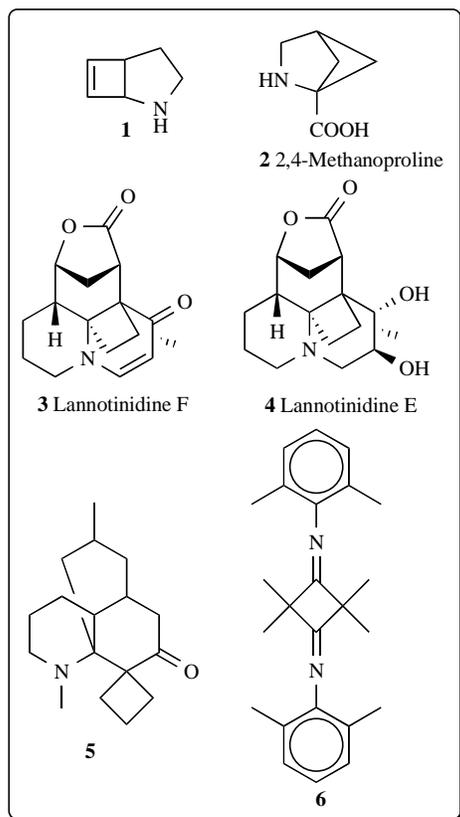
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step consists of a reversible addition of hydrogen cyanide onto the imines (Scheme 1).



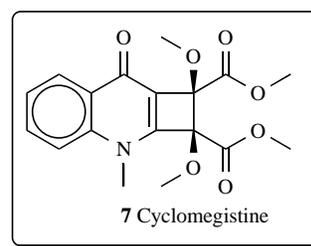
Scheme 1.

Seven *Lycopodium* alkaloids, lannotinidines A-G, have been isolated from the club moss *Lycopodium annotinum* and *L. annotinum* var. *acrifolium*. Two lannotinidines F (**3**) and E (**4**) contain cyclobutane ring elevated NGF mRNA expression. Effects of lannotinidines A–F on neurotrophic factor biosynthesis in 1321N1 human astrocytoma cells were examined by determining NGF mRNA expression. 1321N1 cells were incubated with 30 $\mu\text{g}/\text{mL}$ each of lannotinidines A–F for 6 h, and the mRNA expressions of NGF in 1321N1 cells were examined by a semiquantitative RT-PCR method. The mRNA expressions for NGF were enhanced by lannotinidines B–E, among which (**3**) was the most potent [21].

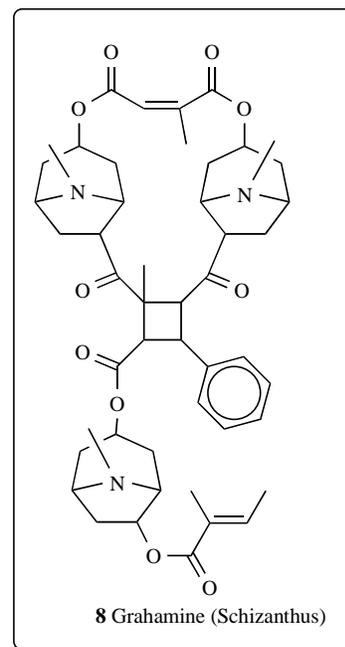


Bioactive hexahydro-1',10'-dimethyl-spiro[cyclo-butane-1,8'(7H)-[1H-5,8a]propane-quinolin]-7'-one (**5**) was isolated from extract of *Lycopodium* species [22]. Unusual tetramethyl-*N,N*-bis(2,6-dimethyl-phenyl)-cyclobutane-1,3-diimine (**6**), triacntanol, and triclin, reported from *Arundo donax*, revealed significant antifeedant activity against the boll weevil, *Anthonomus grand* [23].

Sarcomelicope megistophylla (Rutaceae) is a small to medium sized tree, 8–12 m high, easily recognized by its pubescent leaves, exceptionally large for the genus (up to 35 cm long). This species is endemic to the region of Néaoua, New Caledonia [24]. A new quinolone, cyclomegistine (**7**), was isolated from the bark of *Sarcomelicope megistophylla*. This alkaloid possesses the cyclobuta[b]quinoline ring system that has not been previously described either from natural or from synthetic origin. Biogenetically, cyclomegistine could arise from the oxidative aromatic ring cleavage of an acridone precursor, followed by photo-isomerization of the resulting butadiene into the isomeric cyclobutane [25].

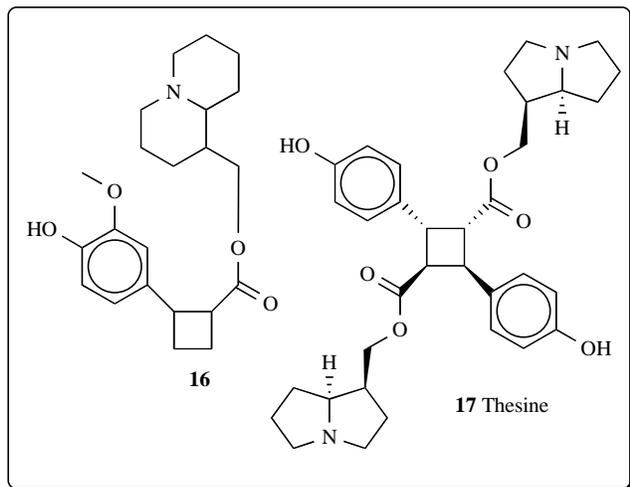


A tropane alkaloid with a 2-methyl-4-phenyl-cyclobutane 1,2,3-tricarboxylic acid ester as the central structure was isolated from the aerial parts of *Schizanthus grahamii* and named grahamine (**8**) [26].

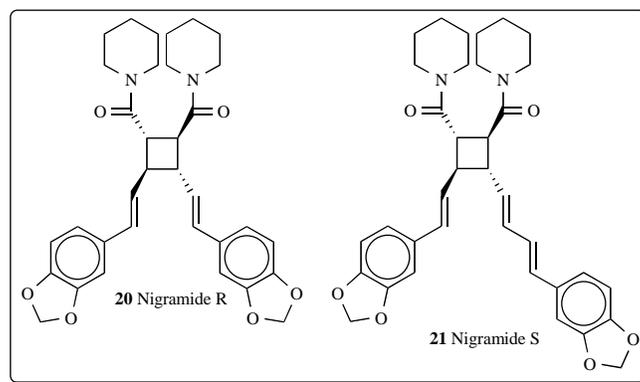
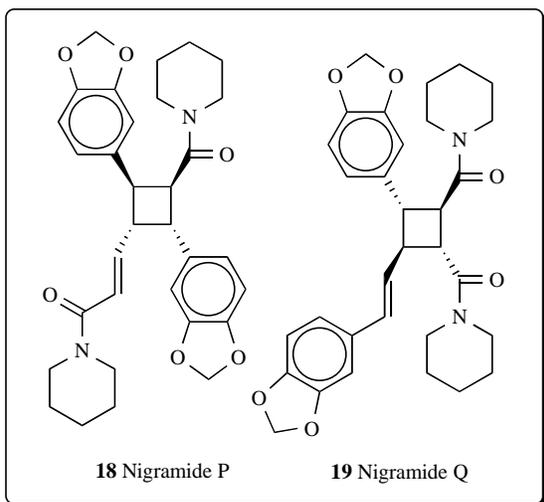


Two new dimeric tropane alkaloids, mooniines A (**9**) and B (**10**) were identified from the leaves of *Erythroxylum moonii* [27].

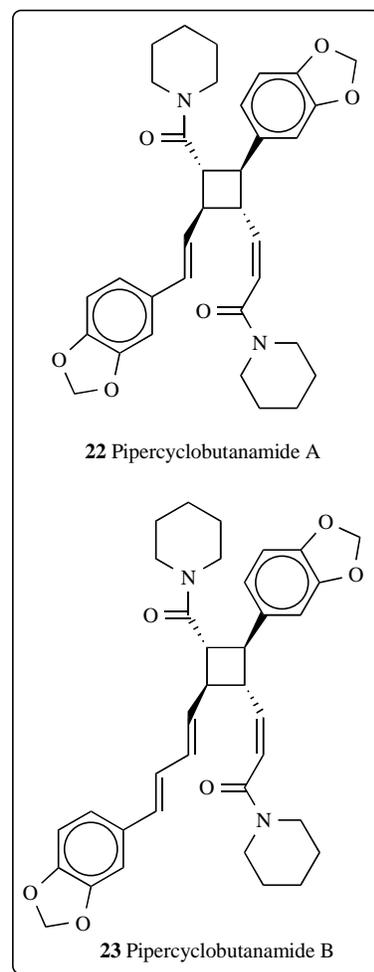
Piper species occurs in the American tropics (700 spp.), followed by Southern Asia (300 spp.), where the economically important species *Piper nigrum* (Black pepper) and *P. betle*



(betel leaf) originated. Patterns of distribution of *Piper* species vary from being locally endemic to widespread. Several cyclobutane-containing alkaloids have been isolated and identified from the genus *Piper* [50]. Fifteen novel dimeric amide alkaloids possessing a cyclohexene ring, nigramides A-O, as well as four novel dimeric amide alkaloids possessing a cyclobutane ring, nigramides P-S (18-21), have been isolated from the roots of *Piper nigrum*. The biosynthetic hypothesis of nigramides A-O was proposed by an intermolecular Diels-Alder reaction from the corresponding monomeric amides [51].



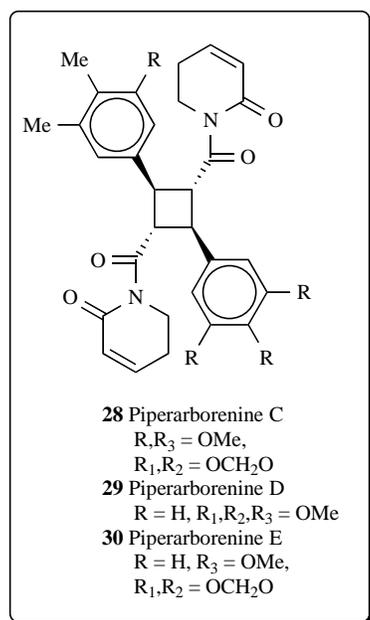
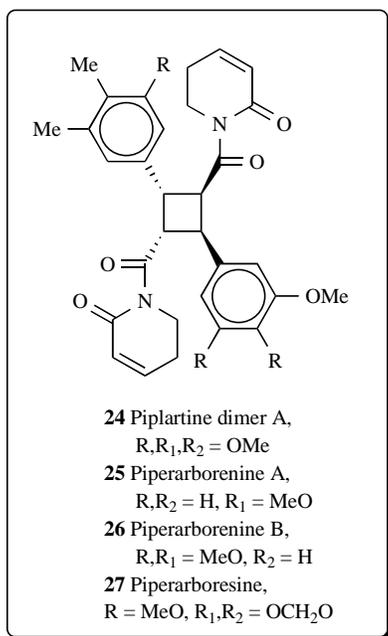
Two other new alkaloids possessing a cyclobutane ring, named pipericyclobutanamides A (22) and B (23), have been isolated from the fruits of *Piper nigrum* [52]. Isolated compounds were potent mechanism-based inhibitors of cytochrome P 450 2D6 (CYP2D6) [53,54].



Two new cyclobutanoid amides, piplartine-dimer A (24), piperarborenine A (25) and B (26), and piperarborensine (27) were isolated from the stem of *Piper arborescens* [55]. Piplartine dimer A (24) has also been obtained from extracts of three piper species: *P. Aborescens*, *P. rugosum*, *P. tuberculatum* [56-58].

Three new cyclobutanoid amides with trans-trans configurations, piperarborenine C (28), piperarborenine D (29)

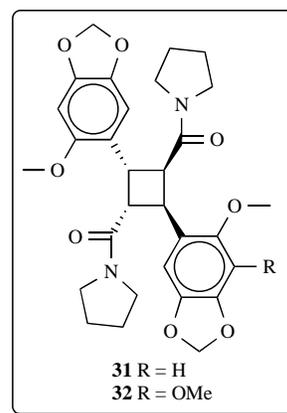
and piperarborenine E (**30**), and a new furanoid lignan, (+)-arborone, together with twelve known compounds, were isolated from the stems of *Piper arborescens*. Piperarborenine C, (+)-diayangamin, pipartine, piperolactam B, piperolactam C, aristolactam BIII, goniotalactam, and methyl *trans*-3,4,5-trimethoxycinnamate possessed antiplatelet aggregation activity *in vitro*. Among them, pipartine showed the most potent anti-platelet aggregation activity induced by collagen and showed an IC₅₀ value of 21.5 μM. Piperarborenines C-E, piperarborenine, aristolactam BIII and goniotalactam showed significant cytotoxic activity (IC₅₀ values < 4 μg/mL) against P-388, HT-29 and A549 cell lines *in vitro* [59].



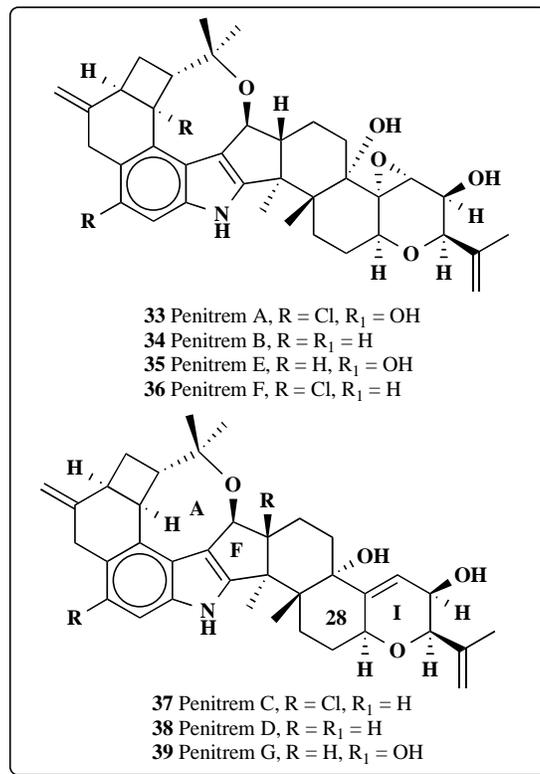
Two new pyrrolidinamide dimers have been isolated from the leaves and stem of *Piper peepuloides* and their

structures established by spectral analysis as cyclobutane-2-(1,3-benzodioxol-5-methoxy-6-yl)-4-(1,3-benzodioxol-4,5-dimethoxy-6-yl)-1,3-dicarboxa-pyrrolidide (**31**) and cyclobutane-2,4-bis-(1,3-benzodioxol-5-methoxy-6-yl)-1,3-dicarboxa-pyrrolidide (**32**) [60].

Tremorgenic mycotoxins produced by fungi belonging to the genera *Penicillium*, *Aspergillus*, and *Claviceps* have potent insecticidal and growth inhibitory activity against the corn earworm (*Heliothis zea*) and the fall armyworm (*Spodoptera frugiperda*) [61-65]. Several similar indole-diterpenoid compounds with cyclobutyl unit have been isolated from these fungi species. Mycotoxins produced by

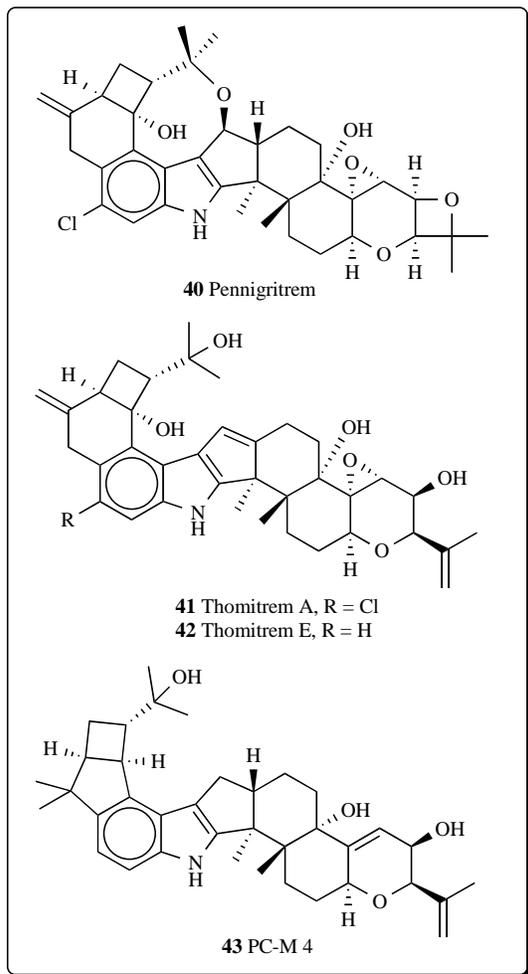


several *Penicillium* species: *P. crustosum*, *P. brevicompactum*, *P. chrysogenum*, *P. expansum*, *P. roqueforti*, *P. spinulosum*, *P. viridicatum*, *P. commune*, *P. citrinum* and *P. solitum*, included mycophenolic acid, roquefortine C, penitrem A-F (**33-38**) and thomitrem A and E (**41** and **42**) [66-74].



Penitrem G (**39**), a new alkaloid, has been isolated together with the already known mycotoxins penitrems A-D and F from the mycelium of *Penicillium crustosum* [75]. Penitrems A-D and F showed convulsive and insecticidal activities against both insect species. A prominent novel analog pennigritrem (**40**) of penitrem A, resolved from the tremorgenic alkaloids of a strain of *Penicillium nigricans*, showed to involve the terminal diterpenoid isoprene in a cyclization which is unique among fungal indole-diterpenoids [76].

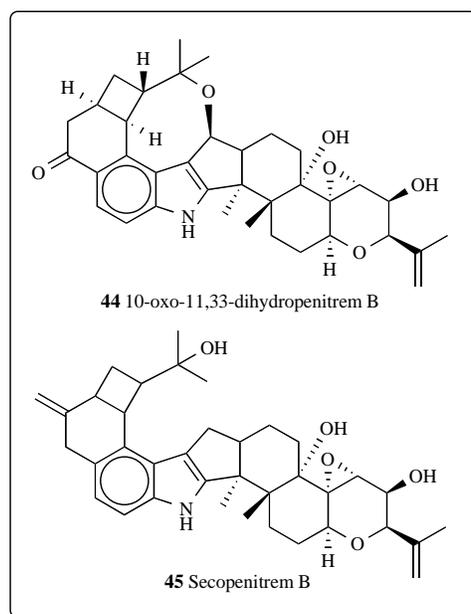
Recently the minor metabolites, PC-M5' and PC-M6, were isolated along with the tremorgenic mycotoxins, penitrems A-F, from the mycelium of *P. crustosum*, were found as contaminants of bread intended for school lunches in Tokyo city. Two new indoloditerpenes, PC-M4 (**43**) and PC-M5, were also isolated from the above fungus. PC-M4 has the same carbon number as the penitrems but a different cyclic ring system. PC-M5 might be a biosynthetic precursor of penitrems and PC-M4 as may also be PC-M5' and PC-M6 [77].



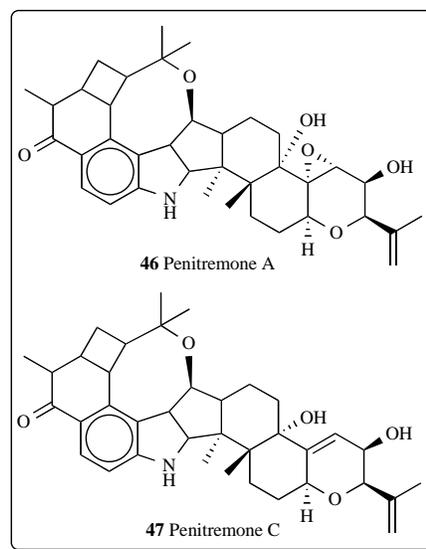
The new antiinsectan metabolite 10-oxo-11,33-dihydropenitrem B (**44**) was isolated from the sclerotia of *Aspergillus sulphureus*. Isolated alkaloid is related to the penitrems, a known group of tremorgenic fungal metabolites. A known aflavinine analog (10,23-dihydro-24,25-dehydroaflavinine) was also isolated from *A. sulphurerus*

sclerotia. This is the first report of any aflavinine analog from a member of the *Aspergillus ochraceus* taxonomic group [78].

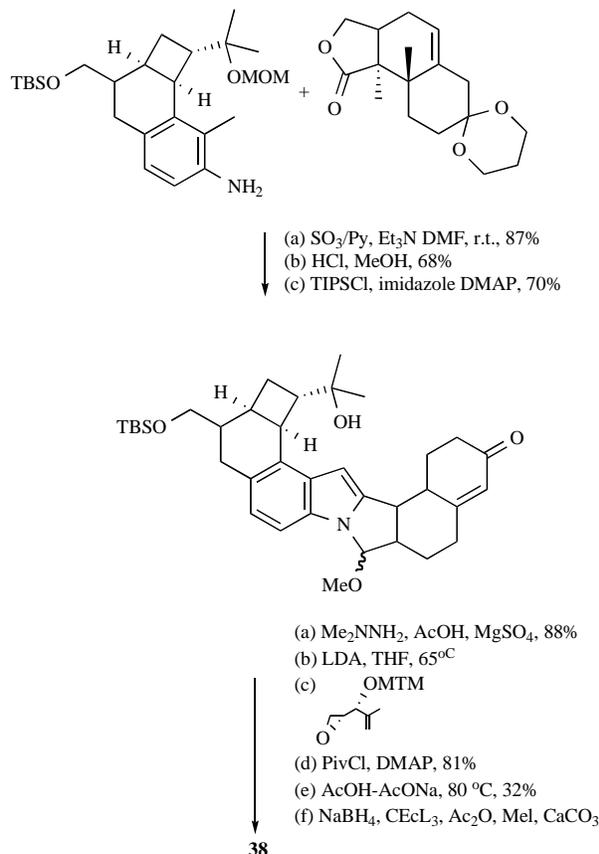
Extract from the sclerotia of *Aspergillus sulphureus* yielded four new antiinsectan compounds of the paspaline/penitrem class, and included secopenitrem B (**45**). Isolated metabolites were structurally related to penitrems. The three indole metabolites, including (**45**), exhibited potent activity against the lepidopteran crop pest *Helicoverpa zea* [79].



A new group of microbial metabolites, designated penitremones A-C, have been characterized by MS and NMR spectroscopy as 10-keto, and 11,33-dihydro-variants of the penitrem indole-isoprenoid skeleton. The principal metabolite penitremone A (**46**), and minor (**47**) produced with penitrem A by a *Penicillium* sp., is an isomer of penitrem E and was also similarly tremorgenic [80]. Different biological activities for penitrem A (**33**) have been studied and reported in some articles [81-88].



Recently, Smith III and co-workers [89] demonstrated the viability of a highly stereoselective tandem Mannich cyclization-grammine fragmentation/addition cascade, critical for assembly of the A and F rings of penitrem D (Scheme 3). They also explored simultaneous execution of this tactic with concurrent construction of ring I. Reinvestigation of a model system provided an explanation for the unanticipated stereochemical outcome at C(28).



Scheme 3.

PREDICTED ACTIVITIES OF ALKALOIDS ISOLATED FROM TERRESTRIAL SPECIES

Probable additional biological activities of cyclobutane-containing alkaloids were evaluated by computer prediction. For this purpose we used computer program 'PASS' [90-92], which predicts about 2,500 pharmacological effects, mechanisms of action, mutagenicity, carcinogenicity, teratogenicity and embryotoxicity on the basis of structural formulae of compounds. PASS predictions are based on structure-activity relationships (SAR) analysis of the training set consisting of about 60,000 drugs, drug-candidates and lead compounds. Algorithm of PASS predictions is described in detail in several publications [91,92]. Using MOL or SD files as an input for the PASS program, user may get a list of probable biological activities for any drug-like molecule as was also published recently [90].

For each activity, P_a and P_i values are calculated, which can be interpreted either as the probabilities of a molecule belonging to the classes of active and inactive compounds,

Table 1. Predicted Biological Activities for Compounds Isolated from Terrestrial Sources

No.	Drug-Likeness	Focal Activity Prediction
1	0.971	$P_a = 0.964$ $P_i = 0.003$ Aryl-acylamidase inhibitor
2	0.991	$P_a = 0.895$ $P_i = 0.006$ Dopamine release stimulant
3	0.965	$P_a = 0.938$ $P_i = 0.005$ Phosphatase inhibitor
4	0.992	$P_a = 0.939$ $P_i = 0.005$ Phosphatase inhibitor
5	0.958	$P_a = 0.840$ $P_i = 0.008$ Membrane integrity antagonist
6	0.055	$P_a = 0.931$ $P_i = 0.001$ D-amino-acid dehydrogenase inhibitor
7	0.832	$P_a = 0.824$ $P_i = 0.007$ Amyotrophic lateral sclerosis treatment
8	0.992	$P_a = 0.728$ $P_i = 0.015$ Dependence treatment
9	0.983	$P_a = 0.769$ $P_i = 0.011$ Dependence treatment
10	0.972	$P_a = 0.820$ $P_i = 0.007$ Dependence treatment
11	0.913	$P_a = 0.775$ $P_i = 0.022$ Neuroprotector
12	0.966	$P_a = 0.855$ $P_i = 0.008$ Pulmonary hypertension treatment
13	0.967	$P_a = 0.848$ $P_i = 0.008$ Pulmonary hypertension treatment
14	0.968	$P_a = 0.841$ $P_i = 0.009$ Pulmonary hypertension treatment
15	0.984	$P_a = 0.794$ $P_i = 0.015$ Pulmonary hypertension treatment
16	0.964	$P_a = 0.838$ $P_i = 0.010$ Dopamine release stimulant
17	0.967	$P_a = 0.808$ $P_i = 0.015$ Dopamine release stimulant
18	0.903	$P_a = 0.937$ $P_i = 0.002$ Cytochrome P450 inhibitor
19	0.889	$P_a = 0.974$ $P_i = 0.001$ Cytochrome P450 inhibitor
20	0.903	$P_a = 0.940$ $P_i = 0.002$ Cytochrome P450 inhibitor
21	0.931	$P_a = 0.960$ $P_i = 0.002$ Cytochrome P450 inhibitor
22	0.954	$P_a = 0.915$ $P_i = 0.003$ Cytochrome P450 inhibitor
23	0.956	$P_a = 0.966$ $P_i = 0.002$ Cytochrome P450 inhibitor
24	0.907	$P_a = 0.919$ $P_i = 0.018$ (-)-(4S)-limonene synthase inhibitor
25	0.882	$P_a = 0.941$ $P_i = 0.020$ (-)-(4S)-limonene synthase inhibitor
26	0.896	$P_a = 0.908$ $P_i = 0.022$ (-)-(4S)-limonene synthase inhibitor
27	0.961	$P_a = 0.853$ $P_i = 0.008$ Pulmonary hypertension treatment
28	0.961	$P_a = 0.853$ $P_i = 0.008$ Pulmonary hypertension treatment
29	0.896	$P_a = 0.908$ $P_i = 0.022$ (-)-(4S)-limonene synthase inhibitor
30	0.950	$P_a = 0.866$ $P_i = 0.038$ (-)-(4S)-limonene synthase inhibitor
31	0.875	$P_a = 0.915$ $P_i = 0.008$ Neuroprotector
32	0.935	$P_a = 0.933$ $P_i = 0.007$ Membrane integrity agonist

respectively, or as the probabilities of the first and second kind of errors in prediction. First kind error of prediction reflects the “false-positives”, when an inactive compound is predicted to be active; and second kind error of prediction: reflects the “false-negatives”, when an active compound is predicted to be inactive.

Interpretation of the predicted results and selection of the most prospective compounds are based on flexible criteria, which depend on the purpose of particular investigation. If the user chooses a rather high value of P_a as a threshold for selection of probable activities, the chance to confirm the predicted activities by the experiment is high too, but many existing activities will be lost. Typically, there are several dozen biological activities in the predicted biological activity spectra; activity that is predicted with the highest probability is called “focal”. Focal biological activities for cyclobutane-containing alkaloids isolated from terrestrial sources are shown below in the Table 1. Additional predicted biological activities for cyclobutane-containing tremorgenic mycotoxins are shown in Table 2.

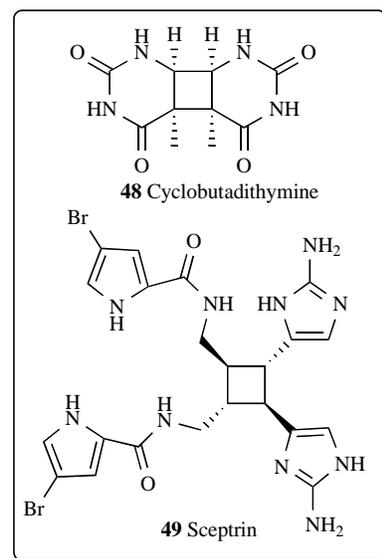
Table 2. Predicted Biological Activities for Tremorgenic Mycotoxins

No.	Drug-Likeness	Focal Activity Prediction
33	0.991	$P_a = 0.736$ $P_i = 0.015$ GABA A receptor antagonist
34	0.993	$P_a = 0.740$ $P_i = 0.014$ GABA A receptor antagonist
35	0.992	$P_a = 0.751$ $P_i = 0.012$ GABA A receptor antagonist
36	0.992	$P_a = 0.724$ $P_i = 0.017$ GABA A receptor antagonist
37	0.993	$P_a = 0.772$ $P_i = 0.056$ Phosphatase inhibitor
38	0.994	$P_a = 0.765$ $P_i = 0.059$ Phosphatase inhibitor
39	0.994	$P_a = 0.760$ $P_i = 0.061$ Phosphatase inhibitor
40	0.988	$P_a = 0.810$ $P_i = 0.038$ Phosphatase inhibitor
41	0.992	$P_a = 0.761$ $P_i = 0.061$ Phosphatase inhibitor
42	0.993	$P_a = 0.734$ $P_i = 0.015$ GABA A receptor antagonist
43	0.993	$P_a = 0.716$ $P_i = 0.025$ Ecdysone 20-monooxygenase inhibitor
44	0.991	$P_a = 0.739$ $P_i = 0.014$ GABA A receptor antagonist
45	0.993	$P_a = 0.742$ $P_i = 0.014$ GABA A receptor antagonist
46	0.993	$P_a = 0.722$ $P_i = 0.018$ GABA A receptor antagonist
47	0.993	$P_a = 0.735$ $P_i = 0.073$ Phosphatase inhibitor

CYCLOBUTANE-CONTAINING ALKALOIDS FROM MARINE SPECIES

Cyclobutane-containing alkaloids have also been isolated and their structures elucidated from marine algae and invertebrates, mainly from marine sponges. Some of them showed different biological activities. The water-soluble cyclobutadithymine (**48**) was extracted from the marine red alga *Porphyra yezoensis*, and showed the protective properties against UV-A irradiation [93].

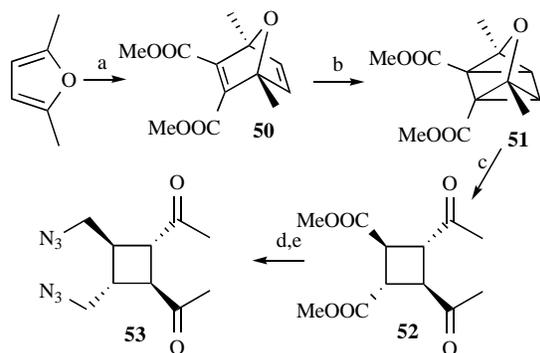
John Faulkner from the University of California (Berkeley) was the first scientist who discovered an antimicrobial agent, scep trin (**49**), from the sponge, *Agelas scep trum*, at 1981. This alkaloid exhibited antimicrobial activity against *Staphylococcus aureus*, *Bacillus subtilis*, *Candida albicans*, *Pseudomonas aeruginosa*, *Alternaria* (fungus), and *Cladosporium cucumerinum*. The compound (**49**) was not toxic at 50 mg/kg [94,95]. The influence of environmental factor, namely depth and sponge-coral interactions as well as the effects of infliction of standardized damages on the production of bromopyrrolic alkaloids isolated from the sponges *Agelas dispar* and *Agelas conifera* were analyzed and scep trin (**49**) was detected in both sponge species [96]. More recently, scep trin and eight dimeric bromopyrrolic alkaloids, nagelamides A-H, and a monomeric one, 9,10-dihydrokeramadine, have been isolated from the Okinawan marine sponge *Agelas* sp. Nagelamides A-H exhibited antibacterial activity against Gram-positive bacteria. Nagelamide G inhibited protein phosphatase 2A activity [97]. Scep trin, ageliferine and xestospongine B, three alkaloids isolated from *Xestospongia* sp. and *Agelas novaecaledoniae* were reported as somatostatin and VIP inhibitors. Scep trin and ageliferine showed an affinity for VIP (19.8 μ M and 19.2 μ M, respectively). Due to the interaction between non-peptidic compounds and somatostatin/VIP receptors, these three alkaloids could be promising agents in the research on natural non-peptidic compounds for therapeutical interventions [98].



The mechanism of action of scep trin was investigated [99]. Scep trin has been reported to exhibit antibacterial and

antifungal activities. Sceptrin demonstrated a bacteriostatic rather than bactericidal effect at the MIC on exponentially growing *Escherichia coli*. Under these conditions, the culture produced chains of cells, and incorporation of radiolabeled precursors into DNA, protein, and cell wall was unaffected, whereas incorporation of ^3H -uridine into RNA was slightly inhibited. At concentrations higher than the MIC, sceptrin was bactericidal, inhibited the incorporation of all radiolabeled precursors, and induced the formation of unusual spheroplasts. Peptidoglycan turnover in *E. coli* appeared to be stimulated by sceptrin, as demonstrated by a release of diaminopimelic acid-containing high mol wt material. Subsequent studies of the release of K^+ from *E. coli* and the lysis of red blood cells suggested that sceptrin disrupts the cell membranes of both prokaryotic and eukaryotic cells. Spheroplast formation may reflect a cell wall effect that occurs subsequent to membrane damage [99]. In a search for potential target sites for C_{11}N_5 compounds obtained from marine sponges of the genus *Agelas*, the authors evaluated their interaction with muscarinic acetylcholine receptors from rat brain membranes. In competition experiments with 3H-QNB, these compounds displayed the following rank order of potency: sceptrin (**49**) > oroidin, \geq dibromosceptrin \geq clathrocin. Sceptrin (**49**) (50 μM) was shown to be a competitive inhibitor of 3H-QNB binding as revealed Scatchard analysis. The results demonstrate the ability of these compounds to interact with multiple molecule targets in the micromolar range [100].

The first enantioselective total synthesis of sceptrin was reported by programming the fragmentation of an oxaquadricyclane [101]. Oxabicyclic (**50**) could be obtained in quantitative yield by Alder reaction. In the [2+2] cyclization to form oxaquadricyclane (**51**), it was found that switching the solvent from diethyl ether to THF not only accelerated the reaction but also avoided potential complications due to the low boiling point of ether (Scheme 4). The highly unstable (**51**) was taken on in crude form to the fragmentation step.



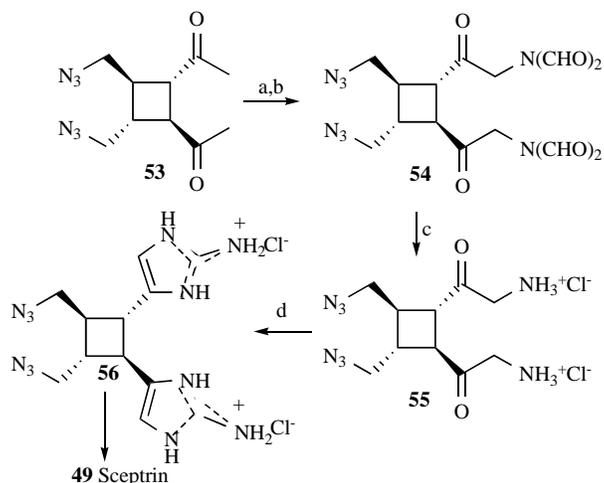
Reagents and conditions: (a) DMAD (1.0 equiv), 2,5-dimethylfuran (1.2 equiv), 1,4-dioxane, 100 °C, 12 h, quantitative; (b) 450 W Hanovia lamp, THF, 24 h, quantitative; (c) H_2SO_4 , MeOH, 24 h, 50%; (d) MeOH, $\text{CH}(\text{OMe})_3$ (14 equiv), TsOH (0.15 equiv), 50 °C, 24 h; DIBAL (6.0 equiv), CH_2Cl_2 , -78 °C, 1.5 h, then AcOH, H_2O , 10 min, quantitative; (e) MsCl (4.4 equiv), 20 °C, 1 h; NaN_3 (6.0 equiv), DMF, 50 °C, 24 h, quantitative.

DMAD = dimethyl acetylenedicarboxylate, THF = tetrahydrofuran, DIBAL = diisobutylaluminum hydride, DMF = *N,N*-dimethylformamide.

Scheme 4.

The known procedure for the fragmentation of oxaquadricyclane (**51**) gave only low yield of cyclobutane (**52**). As this was clearly not a viable option to support extensive synthetic efforts, a more expedient procedure was needed. It was found that the addition of diethyl ether after evaporation of methanol would cause the precipitation of cyclobutane (**52**) in ca. 50% yield, allowing expedient access to multigram quantities of (**52**). With the all-*trans* cyclobutane framework in place, (**53**) can be obtained.

Ketalization of (**56**) under standard conditions (Scheme 5), followed by bromination of the crude ketal with phenyltrimethyl ammonium tribromide, gave bromoketone, displacement of which gave with sodium diformamide compound (**54**), which was hydrolyzed with aqueous HCl to give (**55**) and reacted with cyanamide to form (**56**) in 80% yield over three steps, thereby completing installation of the 2-aminoimidazole, along with 5-10% of an oxazole byproduct [101].



Reagents and conditions: (a) MeOH, $\text{CH}(\text{OMe})_3$ (14 equiv), TsOH (0.15 equiv), 50 °C, 24 h; PTT (2.1 equiv), THF/MeOH/ $\text{CH}(\text{OMe})_3$ 2:1:0.1, 20 °C, 2 h; AcOH(aq), 50 °C, 12 h, 92%; (b) $\text{NaN}(\text{CHO})_2$ (6 equiv), MeCN, 18 h, 20 °C; (c) MeOH, HCl (1% v/v), 16 h; (d) H_2NCN (80 equiv), H_2O , 140 °C microwave, 3 min, 80%.

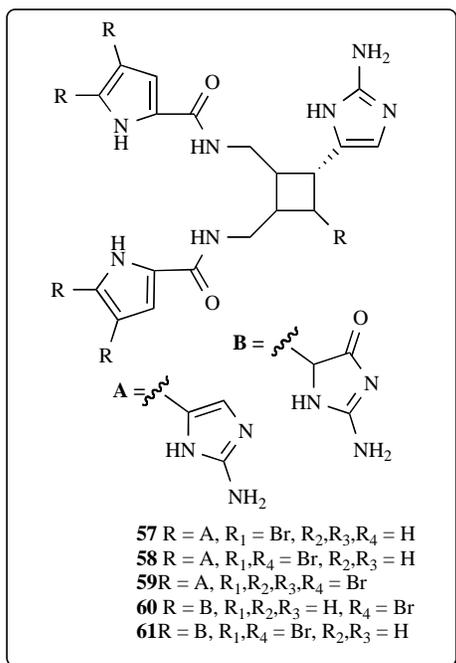
PTT = phenyltrimethylammonium tribromide, THF = tetrahydrofuran.

Scheme 5.

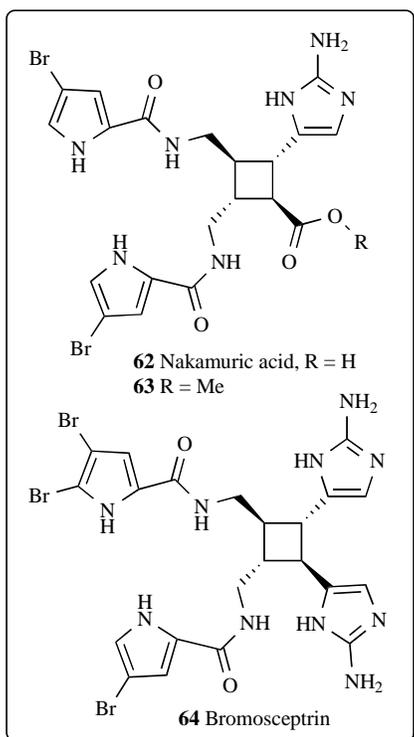
Biologically active extracts of the Caribbean sponge *Agelas conifera* had yielded, in exhaustive studies, the diacetate salts of seven new bromopyrroles (**57-61**), as well as that of the known debromooroidin dimer sceptrin. These compounds were found to be antiviral and antibacterial and were active in barnacle settlement and biochemical prophage induction assays. The oxysceptrins are characterized by their aminoimidazolinone group, the ageliferrins, by a unique cyclohexene-based skeleton [102]. Debromosceptrin (**57**), and two known pyrrole analogs (**58** and **59**) were also found in extract of the Caribbean sponge *Agelas conifera* collected from Belize [103]. Oxysceptrin (**61**) from the Okinawan marine sponge *Agelas cf. nemoechinata* showed active properties against actomyosin ATP-ase [104].

Nakamuric acid (**62**) and sceptrin were identified from Australian sponges belonging to the genus *Agelas* [105]. Two dimeric bromopyrrole alkaloids, nakamuric acid (**62**)

and its corresponding Me ester (**63**), were isolated from the Indopacific sponge *Agelas nakamurai* along with the known



metabolites sceptrin, debromosceptrin, and ageliferin. All compounds inhibited the growth of several Gram-positive and Gram-negative bacteria in the agar plate diffusion assay [106]. Six dimeric bromopyrrole alkaloids were isolated from a Florida Keys specimen of *Agelas conifera*. One of the constituents was identified as a new bromopyrrole metabolite, bromosceptrin (**64**) [107].



PREDICTED ACTIVITIES FOR MARINE ALKALOIDS

Additional predicted activities for natural cyclobutane-containing alkaloids are shown in Table 3.

Table 3. Predicted Biological Activities for Compounds Isolated from Marine Sources

No.	Drug-Likeness	Focal Activity Prediction
48	0.704	P _a = 0.916, P _i = 0.005 Antiepileptic
49	0.965	P _a = 0.669 P _i = 0.009 Prostaglandin E1 antagonist
57	0.899	P _a = 0.682 P _i = 0.008 Prostaglandin E1 antagonist
58	0.914	P _a = 0.719 P _i = 0.007 Prostaglandin E1 antagonist
59	0.921	P _a = 0.719 P _i = 0.007 Prostaglandin E1 antagonist
60	0.896	P _a = 0.616 P _i = 0.013 Prostaglandin E1 antagonist
61	0.908	P _a = 0.625 P _i = 0.010 Prostaglandin E1 antagonist
62	0.880	P _a = 0.672 P _i = 0.009 Prostaglandin E1 antagonist
63	0.837	P _a = 0.631 P _i = 0.012 Prostaglandin E1 antagonist
64	0.927	P _a = 0.682 P _i = 0.008 Prostaglandin E1 antagonist

CONCLUDING REMARKS

Cyclobutane-containing alkaloids are rare group of natural products. They are mainly synthesized by different plant species, and also were detected in some marine species. A little information is known about biological activities of these metabolites. Nevertheless, reported activities for isolated compounds revealed strong antibacterial, antimicrobial, antifeedant, antinociceptive, insecticidal and others activities. The widest spectra of pharmacological activities are exhibited by isolated alkaloids, and/or their N-oxides. Using the program PASS we showed that many reported activities for cyclobutane-containing alkaloids have been predicted, including some additional biological activities.

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