

# Monitoring by HPLC of Chamomile Flavonoids Exposed to Rat Liver Microsomal Metabolism

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**Abstract:** Three major flavonoid chamomile components (quercetin, apigenin-7-O-glucoside and rutin) were subjected to oxidative metabolism by cytochrome P-450 of rat liver microsomal preparations. Changes over time in their respective concentrations were followed using reversed-phase HPLC with UV detection. No clean-up had to be applied as only the specific flavonoid had to be separated from the background components originating from the rat liver microsomes.

Neither the concentration of apigenin-7-O-glucoside nor that of the diglycoside rutin decreased during one hour of exposure to rat microsomal treatment. In contrast, the concentration of quercetin, a lipophilic aglycon, decreased.

Our analytical HPLC results complement the *in silico* calculated lipophilicity (logP) of these compounds; the relatively high lipophilicity of quercetin appears to predispose it to oxidative metabolism in order to decrease its fat solubility. In contrast the much less lipophilic compounds apigenin-7-O-glucoside and rutin were resistant *in vitro* to microsomal treatment.

**Keywords:** Microsomal treatment, cytochrome P-450 dependent oxidation, HPLC, quercetin, apigenin-7-O-glucoside, rutin.

## INTRODUCTION

Chamomile is one of the oldest and best known favourites amongst medicinal herbs and its position as a therapeutic plant with multiple uses is well established. The positive effects derived from chamomile use are related to the presence of its numerous flavonoid constituents. Regarding their core structure many of these constituents are either flavone- (apigenin, luteolin) or flavonol- derivatives (quercetin, patuletin, isorhamnetin) [1]. They appear in various forms such as aglyca, mono- and diglycosides, and/or acyl-derivatives [2].

Some of these constituents have been directly associated with specific therapeutic effects; luteolin and the quercetin aglyca for instance have demonstrated protective and delaying effects on the development of diabetic complications [3].

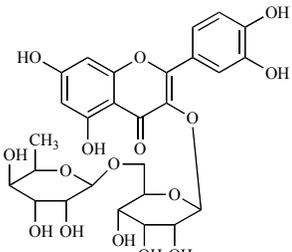
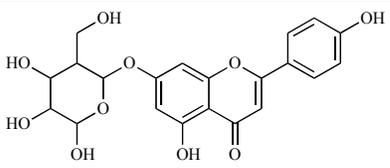
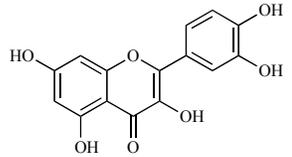
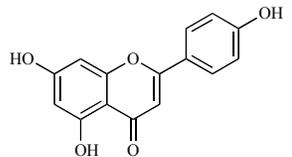
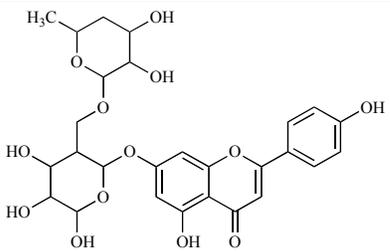
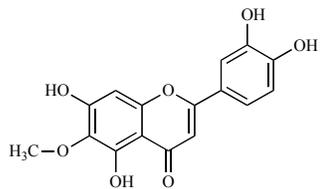
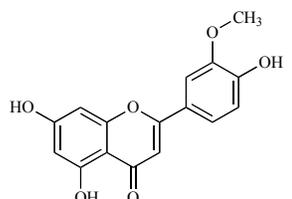
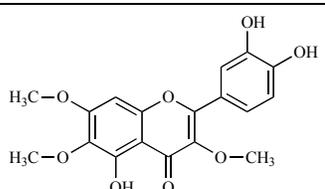
Furthermore apigenin and its -7-O-glucoside derivative are among the most important therapeutic flavonoids; their spasmolytic and antiphlogistic effects are especially prominent [4], in addition to anti proliferative and apoptotic effects in various human cancer cell lines [5].

The presence of apigenin in chamomile is however a matter of some debate as its presence has not been proved unequivocally. Carle *et al.* [6] and Schreiber *et al.* [7] compared extracts derived from freshly collected wild plants with those obtained from greenhouse plants. They concluded that apigenin is not originally present in fresh chamomile, but the result of secondary enzymatic processes. This would explain the great differences in the reported ratios of apigenin and apigenin-7-O-glycoside in the literature, differences which might also be due to different harvesting methods and storage conditions used. Repčák and Martonfi [8] observed that the content of apigenin aglyca in various parts of the flower increases by polyploidization. In di- and tetraploid samples the amount of apigenin aglyca increased by about 15 %. According to both the European and Hungarian Pharmacopoeias [9] the content of apigenin-7-O-glucoside in the Matricaria flower should be at least 0,25 %. This content is measured by HPLC. The HPLC method is suitable for the determination of other flavones too, irrespective of their state of glycosylation and/or acylation [9-15].

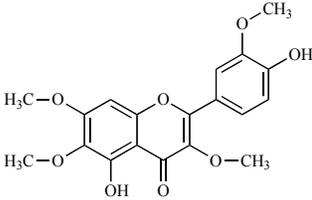
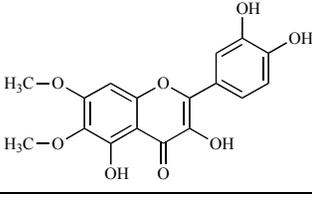
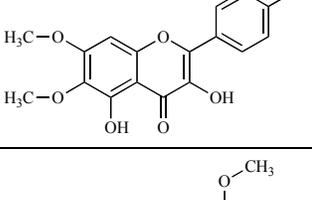
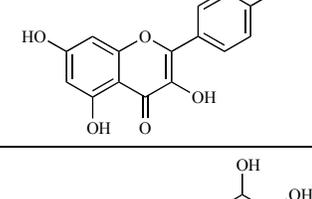
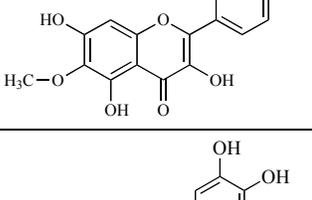
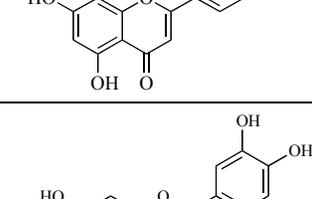
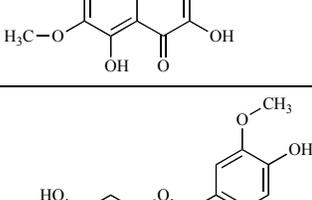
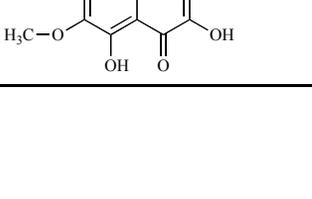
Our aim was to establish whether any of the major matricaria components [quercetin, apigenin-7-O-glucoside and rutin] is subjected to hepatic oxidative microsomal metabolism.

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**Table 1. Name, Chemical Structure, Molecular Size and Composition Calculated Value of Lipophilicity (logP) and Total Polar Surface Area (TPSA, in Angström) of Certain Flavonoids Derived from Chamomile**

#	Name	Structure	MW	logP	TPSA
01	Rutin		C <sub>28</sub> H <sub>10</sub> O <sub>16</sub> 610.57	0.68	265.52
02	Apigenin-7-O-glucoside		C <sub>21</sub> H <sub>20</sub> O <sub>10</sub> 432.41	-0.01	166.14
03	Quercetin		C <sub>15</sub> H <sub>10</sub> O <sub>7</sub> 302.25	1.75	127.45
04	Apigenin		C <sub>15</sub> H <sub>10</sub> O <sub>5</sub> 270.25	2.79	86.99
05	Apigenin-7-O-neohesperidosid		C <sub>27</sub> H <sub>30</sub> O <sub>13</sub> 562.57	0.68	204.83
06	Axillarin		C <sub>16</sub> H <sub>12</sub> O <sub>7</sub> 316.28	2.49	116.45
07	Chrysoeriol		C <sub>16</sub> H <sub>12</sub> O <sub>6</sub> 300.28	2.71	96.22
08	Chrysopenol		C <sub>18</sub> H <sub>16</sub> O <sub>8</sub> 360.34	2.53	114.68

(Table 1) contd.....

#	Name	Structure	MW	logP	TPSA
09	Chrysopenetin		C <sub>18</sub> H <sub>19</sub> O <sub>8</sub> 374.34	2.80	103.68
10	Eupatoletin		C <sub>17</sub> H <sub>14</sub> O <sub>8</sub> 346.31	1.87	125.68
11	Eupaletin		C <sub>17</sub> H <sub>14</sub> O <sub>7</sub> 330.31	2.42	105.45
12	Isorhamnetin		C <sub>16</sub> H <sub>12</sub> O <sub>7</sub> 316.28	2.05	116.45
13	Jaceidin		C <sub>18</sub> H <sub>16</sub> O <sub>8</sub> 360.34	2.53	114.68
14	Luteolin		C <sub>15</sub> H <sub>10</sub> O <sub>8</sub> 286.25	2.39	107.22
15	Patuletin		C <sub>16</sub> H <sub>12</sub> O <sub>8</sub> 332.28	1.64	136.68
16	Spinacetin		C <sub>17</sub> H <sub>14</sub> O <sub>8</sub> 346.31	1.87	125.68

## EXPERIMENTAL

**Materials:** HPLC grade acetonitrile, methanol and trifluoroacetic acid were purchased from Merck (Darmstadt, Germany). Water was double-distilled and deionized. Rutin, apigenin-7-O-glucoside and quercetin were purchased from Sigma-Aldrich (St. Louis, MO, USA). The chemical structures of rutin, apigenin-7-O-glucoside and quercetin are shown in Table 1. The Zorbax RX-C-18 column and precolumn were purchased from Kromat Kft. (Budapest, Hungary).

**Microsomal Preparation:** Quercetin, apigenin-7-O-glucoside and rutin (25  $\mu\text{M}$ , each) were incubated separately for 0, 15, 30, 45 and 60 minutes with rat liver microsomes (Gedeon Richter Plc., Budapest, Hungary, Lot no. 081022, 0.5  $\text{mg mL}^{-1}$ ) in 4 mL incubation medium also consisting of Tris-HCl buffer (0.12 mM, pH 7.4 at 37°C),  $\text{MgCl}_2$  (5 mM), sodium phosphate (6.25 mM), D-glucose-6-phosphate (5 mM), D-glucose 6-phosphate dehydrogenase (1  $\text{U mL}^{-1}$ ), and the reduced form of nicotinamide adenine dinucleotide phosphate (NADPH, 0.5 mM). Parallel experiments were carried out using each substance (quercetin, apigenin-7-O-glucoside, rutin) in the buffered medium but without any D-glucose-6-phosphate, D-glucose 6-phosphate dehydrogenase and the reduced form of nicotinamide adenine dinucleotide phosphate. A control experiment that subjected the entire mixture to microsomal treatment (rat liver microsomes, in the solution of Tris-HCl buffer at pH 7.4,  $\text{MgCl}_2$ , sodium phosphate, D-glucose-6-phosphate, D-glucose 6-phosphate dehydrogenase, and the reduced form of nicotinamide adenine dinucleotide phosphate) but without any substrate to be metabolized was also performed. Microsomal treatment was done using a rotating, thermostat-regulated water bath at 37°C. Incubation with microsomal preparation was terminated after 0, 15, 30 and 60 min respectively, by adding a 4-fold excess of methanol. Each experiment was performed in duplicate. Incubated samples were placed at -20 °C for 10

minutes, and centrifuged using 2500 x g at 4 °C (Sigma Laborzentrifugen, Osterode am Harz, Germany) and finally stored at -80 °C before their reversed-phase HPLC measurement.

**Chromatography:** A JASCO made HPLC system was used that contained a solvent delivery system (PU 1580 Pump), degasser (DG-2080-54), an autoinjector (AS-2057 Plus), and a UV detector (UV-1575). Elution of rutin, apigenin-7-O-glucoside and quercetin was monitored at 257 nm.

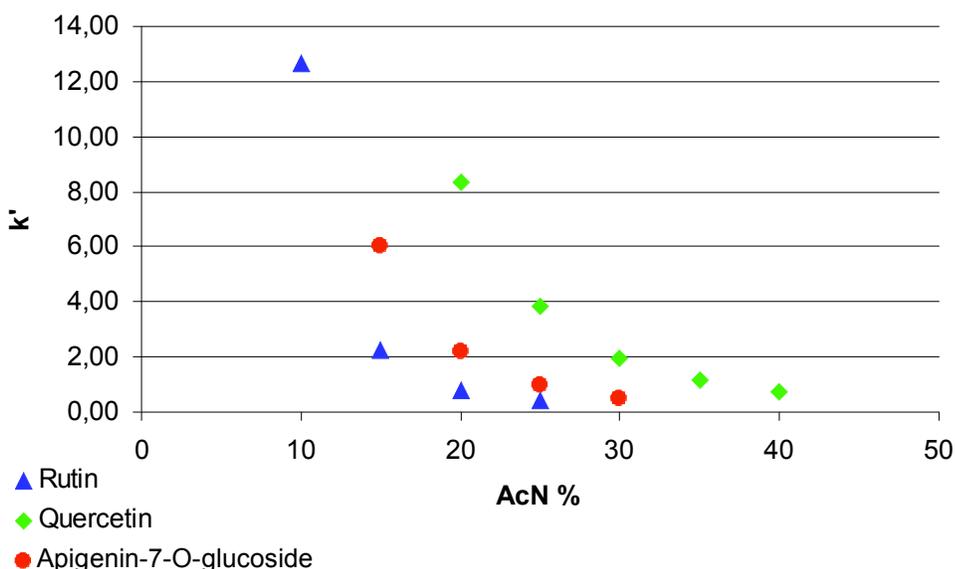
The samples were not subjected to any clean-up. Fifty  $\mu\text{L}$  of the samples were injected. The separation was done on a 250 mm x 4.6 mm I.D., 5- $\mu\text{m}$  particle size Zorbax RX-C-18 column protected by a 12.5 mm x 4.6 mm, I.D. precolumn containing the same packing material. The mobile phases were acetonitrile—water 40:60, 35:65, 30:70, 25:75, 20:80, 15:85 and 10:90, and each mobile phase contained 0.1% of trifluoroacetic acid (TFA). The mobile phase flow rate was 1  $\text{mL min}^{-1}$ , the separations were done at ambient temperature (24°C  $\pm$  1°C). Chromatograms were electronically stored and evaluated by the use of Borwin 1.50 chromatography software (JMBS, Le Fontanil, France).

**Calibration:** Calibrations of the substances were done covering 0.5  $\mu\text{g mL}^{-1}$  through 20  $\mu\text{g mL}^{-1}$  range for quercetin, apigenin-7-O-glucoside and rutin.

**Lipophilicity Calculations:** The logP value, and the total polar surface area (TPSA) were calculated using the Pallas program of CompuDrug [16,17].

## RESULTS

The physico-chemical parameters of several chamomile flavonoids belonging to three major groups [aglycons (flavonoid compounds without any substituting sugar), monoglycosylated and diglycosylated] were calculated using the Pallas program. Chemical structures, molecular



**Fig. (1).** The elution of quercetin, apigenin-7-O-glucoside and rutin are characterized by their  $k'$  values when using various compositions of the mobile phases.

composition, molecular size, logP and total polar surface area (TPSA in Angström) are given in Table 1. Flavonoid aglycons were rather lipophilic with a logP value higher than 1.5.

The mobile phase composition for HPLC of quercetin, apigenin-7-O-glucoside and rutin was optimized to avoid any interference with the background signal of the microsomal incubation medium. Different ratios of the organic modifier and water in the mobile phases were used, such as acetonitrile—water 50:50, 45:55, 40:60, 35:65, 30:70, 25:75, 20:80, 15:85, 10:90 (Fig. 1). Using the optimum mobile phase compositions, the peaks of quercetin, apigenin-7-O-glucoside and rutin eluted at 7.10, 7.72 and 7.73 minutes with the ratio of acetonitrile and water (30:70) for separation of quercetin, apigenin-7-O-glucoside (20:80) and rutin (15:85), respectively. (Fig 2, from left to right).

No interference of any one of the flavonoid compound peaks with those from the microsomal medium was found. Fig. (3) shows that calibration for all substances produced a straight line characterized by their respective parameters,  $a$ ,  $y_0$  and  $R^2$  (from the equation of the straight line:  $y = ax + y_0$ ), as shown in Table 2.

Molecular size (MW), lipophilicity (logP) and total polar surface area (TPSA) of several flavonoids were calculated using the Pallas Program of CompuDrug Inc. These values are given in Table 1.

## CONCLUSIONS

Chamomile [the dried flower heads of *Matricaria recutita* L. (*Chamomilla recutita* [L.] Reuschert)] has been used for centuries as a home remedy; its therapeutic value is well established as documented by the inclusion in numerous Pharmacopoeias.

In traditional medicine the plant is mostly used as chamomile tea, which is administered p.o. for alleviation of gastro-intestinal complaints [18]. Externally chamomile is applied on skin and/or mucous membranes at sites of minor inflammation, including the oral cavity and the gums (mouthwashes), the respiratory tract (inhalations), and the anal and genital areas (baths, ointments) [19,20].

The main constituents of chamomile are the essential oil and the flavone derivatives [21-23].

*In vitro* liver microsomal treatment of organic compounds serves as a model for the first pass effect of the

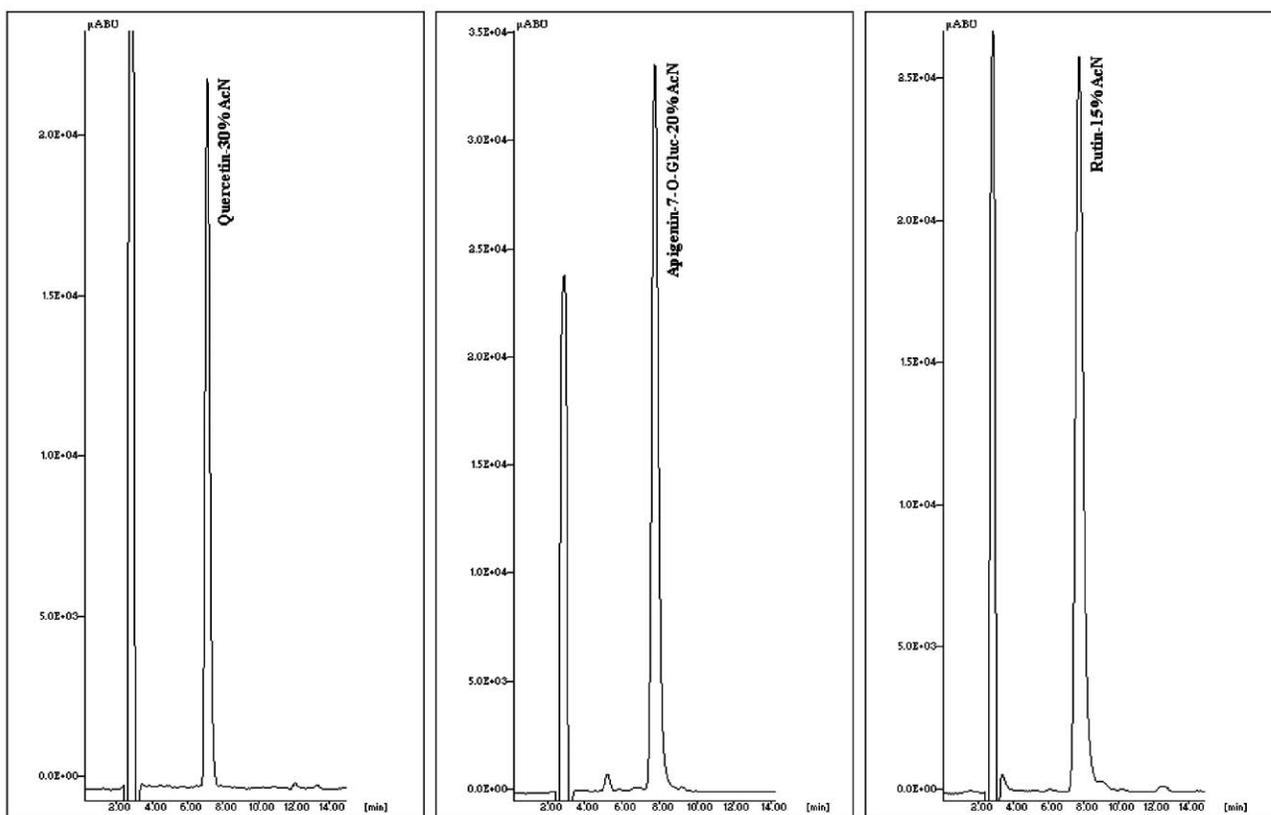


Fig. (2). Typical chromatograms of the microsomal medium spiked with the appropriate amount of quercetin, apigenin-7-O-glucoside and rutin in panels a, b and c respectively.

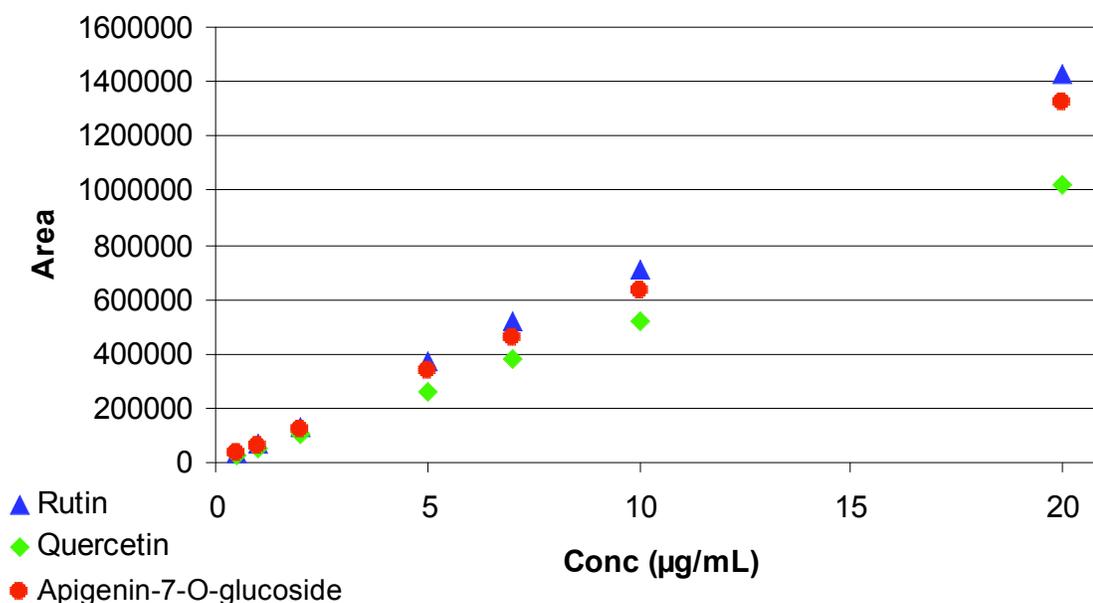


Fig. (3). Calibration curve for quercetin, apigenin-7-O-glucoside and rutin using the adequate mobile phase.

Table 2. Characteristics of the Calibration Lines ( $y = ax + y_0$ ) of RP-HPLC Quantitative Determination of Quercetin, Apigenin-7-O-Glucoside and Rutin

Compound	a (Slope)	$y_0$	$R^2$
Quercetin (03)	51077	3701	0.9993
Apigenin-7-O-glucoside (02)	66236	8101	0.9991
Rutin (01)	71362	2246	0.9995

liver in a living animal. Information may be obtained both on the putative fate of the compound in the body, and also the preferred way of administration. Liver microsomes generally metabolize xenobiotics (foreign compounds ingested) using oxidative metabolism *via* the cytochrome P-450 system. The lipophilicity of xenobiotics decreased *via* these reactions.

Fig. (4) compares the relative changes of the concentrations of quercetin, apigenin-7-O-glucoside and rutin during microsomal treatment. While the concentration of rutin and that of apigenin-7-O-glucoside remain unchanged, that of quercetin decreases.

The most lipophilic flavonoid, the aglycon quercetin undergoes cytochrome P-450-dependent microsomal

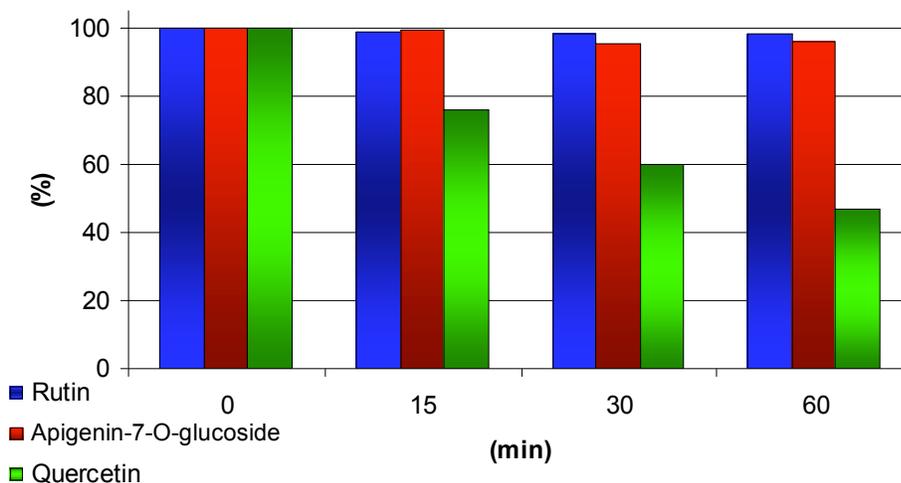


Fig. (4). Concentration of quercetin, apigenin-7-glucoside and rutin through the microsomal treatment at 0, 15, 30 and 60 min of the treatment.

metabolism, while the two less lipophilic compounds examined (apigenin-7-O-glucoside and rutin) were resistant to the effect of rat liver microsome (Fig. 4).

## REFERENCES

- [1] Carle, R.; Isaac, Q. Fortschritte in der Kamillenforschung in den Jahren 1974 bis 1984. *Dtsch. Apoth. Ztg.*, **1985**, *125*(Suppl 1), 2-8.
- [2] Švehliková, V.; Bennett, R.N.; Mellon, F.A.; Needs, P.W.; Piacente, S.; Kroon, P.A.; Bao, Z. Isolation, identification and stability of acylated derivatives of apigenin 7-O-glucoside from chamomile (*Chamomilla recutita* [L.] Rauschert). *Phytochemistry*, **2004**, *65*, 2323-2332.
- [3] Kato, A.; Minoshima, Y.; Yamamoto, J.; Adachi, I.; Watson, A.A.; Nash, R.J. Protective effects of dietary chamomile tea on diabetic complications. *J. Agricult. Food Chem.*, **2008**, *56*, 8206-8211.
- [4] Petri, G. *Gyógynövények és készítményeik a terápiában* (Medicinal Plants and their Products in Therapy, in Hungarian), Galenus Publisher: Budapest, **2006**.
- [5] Srivastava, J.K.; Gupta, S. Anti proliferative and apoptotic effects of chamomile extract in various human cancer cells. *J. Agricult Food Chem.*, **2007**, *55*, 9470-9478.
- [6] Carle, R.; Dölle, B.; Müller, W.; Baummeister, U. Thermospray liquid-chromatography mass-spectrometry (tsp lc/ms) - analysis of acetylated apigenin-7-glucosides from *chamomilla-recutita*. *Pharmazie*, **1993**, *48*, 304-306.
- [7] Schreiber, A.; Carle, R.; Reinhard, E. On the accumulation of apigenin in chamomile flowers. *Planta Med.*, **1991**, *56*, 179-181.
- [8] Repčák, M.; Martonfi, P. The variability pattern of apigenin glucosides in *chamomilla-recutita* diploid and tetraploid cultivars. *Pharmazie*, **1995**, *50*, 696-699.
- [9] *Pharmacopoea Hungarica*, Ed. VIII. Tomus II., Medicina, Budapest, **2004**, pp. 1944-1946, 2203-5.
- [10] Carle, R. In *Hagers Handbuch der Pharmazeutischen Praxis*, 5th ed., Hänsel, R.; Keller, K.; Rimpler, H.; Schneider, G., Eds.; Springer: Berlin, **1992**, *4*, pp. 817-831.
- [11] Kakasy, A.; Marczal, G.; Héthelyi, É.; Lemberkovic, É. Dracocephalum fajok mikromorfológiai és fitokémiai jellemzése (Micromorphology and phytochemistry of certain Dracocephalum species, in Hungarian). *Botanikai Közlemények*, **2004**, *91*, 140.
- [12] Papp, I.; Apáti, P.; Andrasek, V.; Blázovics, A.; Balázs, A.; Kursinszki, L.; Kite, G.C.; Houghton, P.J.; Kéry, Á. LC-MS analysis of antioxidant plant phenoloids. *Chromatographia*, **2004**, *60*, S93-S100.
- [13] Kakasy, A.; Füzfai, Z.; Kursinszki, L.; Molnár-Perl, I.; Lemberkovic, É. Analysis of non-volatile constituents in Dracocephalum species by HPLC and GC-MS. *Chromatographia*, **2006**, *63*, S17-S22.
- [14] Weber, B.; Herrmann, M.; Hartmann, B.; Joppe, H.; Schmidt, C.O.; Bertram, H.J. HPLC/MS and HPLC/NMR as hyphenated techniques for accelerated characterization of the main constituents in Chamomile (*Chamomilla recutita* [L.] Rauschert). *Eur. Food Res. Technol.*, **2008**, *226*, 755-760.
- [15] Szőke, É.; Kéry, Á.; Lemberkovic, É. *Schweiz Pharmacognosy. Pharmacobotanical and Phytochemical Investigations of Plant Drugs*. Semmelweis Kiadó, Budapest, **2009**, (in press).
- [16] Pallas Program: www.compudrug.com
- [17] Molnár, L.; Keserű, G.M.; Papp, Á.; Gulyás, Z.; Darvas, F. A neural network based classification scheme for cytotoxicity predictions: validation on 30,000 compounds. *Bioorg. Med. Chem. Lett.*, **2006**, *16*, 81037-81039.
- [18] Franke, R.; Schilcher, H. *Chamomile Industrial Profiles*, CRC Press: Boca Raton, **2005**.
- [19] Carle, R.; Isaac, O. Die Kamille – Wirkung und Wirksamkeit. *Z. Phytotherapie*, **1987**, *8*, 67.
- [20] Förster, C.F.; Süssmann, H.E.; Patzelt-Wenzler, R. *Schweiz. Rundschau Med.*, **1996**, *85*, 1476.
- [21] Schilcher, H. *Die Kamilla – Handbuch für Ärzte, Apotheker und anderen Naturwissenschaftler*, Wiss. Verlagsges, Stuttgart, Germany, **1987**, p. 152.
- [22] Szőke, É.; Máday, E.; Tyihák, E.; Kuzovkina, I.N.; Lemberkovic, É. New terpenoids in cultivated and wild chamomile (*in vivo* and *in vitro*). *J. Chromatogr. B*, **2004**, *800*, 231-238.
- [23] Shikov, A.N.; Laakso, I.; Pozharitskaya, O.N.; Darman, H.J.D.; Makarov, V.G.; Tikhonov, V.P.; Hiltunen, R. Identification of spiroketal polyacetylenes as the main components of an oil extract of chamomile (*Chamomilla recutita* L. Rausch.) flowers. *Planta Med.*, **2006**, *72*, 1026.

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